

## New Diterpenoids from *Tinospora capillipes*

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A phytochemical study of the roots of *Tinospora capillipes* resulted in the characterization of two new highly unsaturated diterpenoids, tinocapilactones A and B (**1** and **2**), as well as of six known ones. The structures of **1** and **2** were assigned on the basis of extensive 1D- and 2D-NMR experiments, combined with mass-spectrometric methods.

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**Introduction.** – Species of the genus *Tinospora*, widely distributed over Asia and Africa, are well-known for their medicinal properties [1]. The distinctive constituents of this genus are the clerodane-based furonoid diterpenoids [2]. *Tinospora capillipes* GAGNEP, a member of the Menispermaceae family, is a special plant native in South China. Its rhizome (*Radix Tinosporae*, *R.T.*) has been used in traditional Chinese medicine (TCM) for treating sore throat, laryngitis, gastralgia, and diarrhea for thousands of years [3]. A series of clerodane-type diterpenes, botanic steroids, and alkaloids isolated from this species were reported [4–8]. In the present study, two new highly unsaturated diterpenoids, namely tinocapilactones A and B<sup>1)</sup> (**1** and **2**), were isolated from the roots of *T. capillipes*, together with six known ones, *i.e.*, columbin (**3**) [9], palmatoside C (**4**) [10], isocolumbin (**5**) [11], tinocallones A and B (**6** and **7**), and epitinophylloside (**8**) [7]. Their structures were established by mass-spectrometric and spectroscopic analyses, especially 2D-NMR techniques (HMQC, HMBC, and NOESY).

**Results and Discussion.** – Tinocapilactone A (**1**) was obtained as an optically active, colorless, amorphous powder. The HR-ESI-MS ( $m/z$  395.1473 ( $[M + Na]^+$ )) revealed a molecular formula  $C_{23}H_{28}O_9$ , suggesting the presence of ten degrees of unsaturation. Strong IR absorption bands at 1746 and 1716  $cm^{-1}$  were attributable to  $\delta$ -lactone and ester CO groups, respectively. The  $^{13}C$ -NMR spectrum indicated the presence of 23 C-atoms comprising four CO groups ( $\delta(C)$  176.2, 175.0, 167.6, and 165.4), three olefinic C-atoms ( $\delta(C)$  147.9, 147.7, 136.8, 129.5, 125.2, and 118.6), an O-bearing CH group ( $\delta(C)$  75.2), one O-bearing quaternary C-atom ( $\delta(C)$  80.0), and three MeO groups ( $\delta(C)$  52.5, 52.0, and 51.7) (*Table*). Among the four CO groups, two relatively high-field C-atom signals ( $\delta(C)$  167.6 and 165.4) were assigned to two  $\alpha,\beta$ -unsaturated ester CO groups, which was confirmed by the UV maximum at 264 nm ( $\log \epsilon$  4.09). The

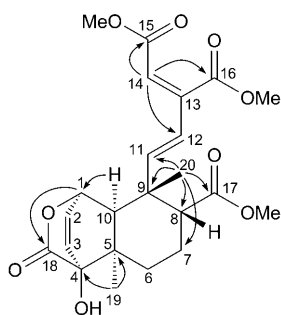
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<sup>1)</sup> Trivial atom numbering; for systematic names, see *Exper. Part*.



Table.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data ( $\text{CDCl}_3$ , 500 and 125 MHz, resp.) of **1** and **2**.  $\delta$  in ppm,  $J$  in Hz.

	<b>1</b>		<b>2</b>	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
H–C(1)	5.20 ( <i>d</i> , $J=4.5$ )	75.2	5.26 ( <i>d</i> , $J=4.3$ )	74.9
H–C(2)	6.42 ( <i>dd</i> , $J=7.7, 5.0$ )	129.5	6.42 ( <i>dd</i> , $J=7.7, 5.2$ )	129.6
H–C(3)	6.32 ( <i>dd</i> , $J=7.7, 1.2$ )	136.8	6.33 ( <i>d</i> , $J=7.7$ )	136.8
C(4)	–	80.0	–	79.8
C(5)	–	37.0	–	36.8
$\text{CH}_2(6)$	1.62–1.65 ( <i>m</i> , $\text{H}_\alpha$ ), 1.55–1.60 ( <i>m</i> , $\text{H}_\beta$ )	27.5	1.64–1.68 ( <i>m</i> , $\text{H}_\alpha$ ), 1.54–1.60 ( <i>m</i> , $\text{H}_\beta$ )	28.1
$\text{CH}_2(7)$	1.85–1.92 ( <i>2m</i> )	20.6	1.84–1.92 ( <i>2m</i> )	20.9
H–C(8)	2.58 ( <i>t</i> , $J=8.5$ )	47.0	2.63 ( <i>t</i> , $J=8.0$ )	46.7
C(9)	–	40.6	–	40.6
H–C(10)	1.82 ( <i>s</i> )	52.6	1.78 ( <i>s</i> )	53.7
H–C(11)	6.44 ( <i>d</i> , $J=16.3$ )	147.9	7.04 ( <i>d</i> , $J=16.4$ )	149.8
H–C(12)	6.12 ( <i>d</i> , $J=16.3$ )	125.2	6.29 ( <i>d</i> , $J=16.4$ )	118.8
C(13)	–	147.7	–	159.1
H–C(14)	5.82 ( <i>s</i> )	118.6	5.91 ( <i>s</i> )	117.4
C(15)	–	165.4	–	170.4
C(16)	–	167.6	5.96 ( <i>s</i> )	102.7
C(17)	–	175.0	–	175.4
C(18)	–	176.2	–	176.1
Me(19)	1.11 ( <i>s</i> )	25.7	1.09 ( <i>s</i> )	25.5
Me(20)	1.35 ( <i>s</i> )	25.9	1.39 ( <i>s</i> )	26.4
MeO–C(15)	3.91 ( <i>s</i> )	52.5	–	–
MeO–C(16)	3.76 ( <i>s</i> )	52.0	3.57 ( <i>s</i> )	56.0
MeO–C(17)	3.63 ( <i>s</i> )	51.7	3.64 ( <i>s</i> )	51.7

Fig. 1. Selected HMBC ( $\text{H} \rightarrow \text{C}$ ) correlations of **1**

group of C(17) must be  $\alpha$ -oriented. The correlation H–C(8)/ $\text{H}_\beta$ –C(6) indicated that  $\text{H}_\beta$ –C(6) was also  $\beta$ -configured. The NOE correlations  $\text{H}_\alpha$ –C(6)/Me(19), Me(19)/H–C(10), and H–C(10)/H–C(1) indicated that H–C(1), H–C(10), and Me(19) are on the  $\alpha$ -face. Complete  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR assignments (Table) were accomplished by a combination of 2D-NMR techniques, including HMQC, HMBC, and NOESY. Thus, the structure of tinocapilactone A<sup>1</sup> (**1**) was established.

Tinocapilactone B (**2**) had the molecular formula  $\text{C}_{22}\text{H}_{26}\text{O}_8$  based on HR-ESI-MS analysis ( $m/z$  419.1710 ( $[\text{M} + \text{H}]^+$ )). The compound was also obtained as colorless, amorphous powder. Its NMR spectra showed similarity with those of **1**, except for the

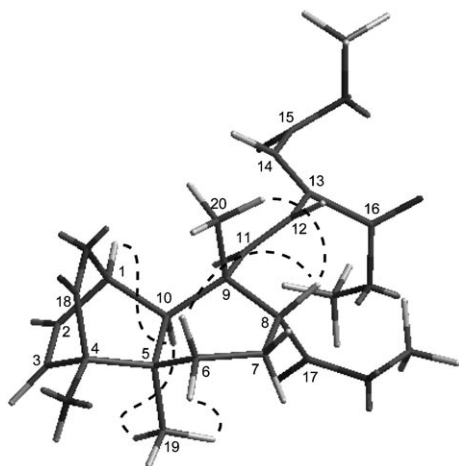


Fig. 2. Key NOESY correlations and relative configuration of **1**

signals of C(11) to C(16), which suggested that **2** possesses the same core (clerodane-type diterpene with  $\delta$ -lactone) as **1** but differs in the side chain. The structure of this side chain was determined to be an ethenyl-linked  $\alpha,\beta$ -unsaturated lactone moiety with a MeO group at C(16) by comparison of the NMR data of **2** with those of yunnanconarin C [12]; this was confirmed by HMBC data (Fig. 3). The interpretation of the  $^1\text{H-NMR}$  and NOESY data revealed that **2** has the same relative configuration as **1**. Thus, the structure of tinocapilactone B<sup>1</sup> (**2**) was established, except for the relative configuration at C(16).

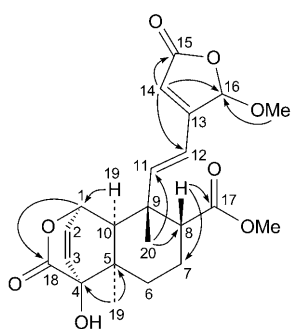


Fig. 3. Selected HMBC (H  $\rightarrow$  C) correlations of **2**

The present study showed that the major constituent, tinocapilactone A (**1**), with significant anti-inflammatory activities may be the bioactive compound of *T. capillipes* [13]. The study also revealed that the constituents of this plant are similar to those of *T. sagittata* (especially concerning the major constituents, such as compounds **3**, **4**, and **8**) [14], which can explain why these two species are used for the same treatments in the traditional folk medicine in China.

### Experimental Part

*General.* All solvents used were of anal. grade (*Shanghai Chemical Plant*). TLC: precoated silica gel *GF<sub>254</sub>* plates (*Qingdao Haiyang Chemical Plant*). Column chromatography (CC): silica gel (SiO<sub>2</sub>; 230–400 mesh), SiO<sub>2</sub> *H-60*, and *MCI-CHP20P* gel (75–150  $\mu$ ; *Mitsubishi Chemical Industries Ltd.*). Optical rotations: *Perkin-Elmer 341* polarimeter. UV Spectra: *Shimadzu UV-2450* spectrometer;  $\lambda_{\max}$  (log  $\epsilon$ ). IR Spectra: *Thermo Nicolet-6700* spectrophotometer; in cm<sup>-1</sup>. NMR spectra: *Bruker AM-500* apparatus;  $\delta$  in ppm rel. to Me<sub>4</sub>Si,  $J$  in Hz. ESI-MS: *Agilent 6210-Lc/Tof* mass spectrometer; in  $m/z$ .

*Plant Material.* The plant material used for this study was collected from Guangxi Province, P. R. China, in August 2007, and identified by Prof. *Yong-Hong Zhang* of the Fujian Medical University, P. R. China. A voucher specimen (No. 20070823T) was deposited with the Zhejiang University of Technology.

*Extraction and Isolation.* The air-dried roots of *T. capillipes* (3.2 kg) were coarsely powdered and percolated with 95% EtOH/H<sub>2</sub>O. After solvent removal, the crude extract (252 g) was suspended in H<sub>2</sub>O (3 l) and extracted with AcOEt (5  $\times$  500 ml) to afford the AcOEt-soluble fraction. The AcOEt-soluble fraction (72 g) was subjected to CC (SiO<sub>2</sub>, petroleum ether/acetone 4:1  $\rightarrow$  0:1, then MeOH): to give three major fractions, *Fr. 1* (43 g), *2* (3.0 g), and *3* (10.1 g). Compound **3** (30 g) was purified by recrystallization of *Fr. 1* from petroleum ether/acetone 2:1. The filtrate was evaporated to give a yellow residue, which was further purified by CC (SiO<sub>2</sub>, petroleum ether/AcOEt 4:1  $\rightarrow$  2:1): **1** (1.6 mg), **2** (6.0 mg), and **5** (50 mg). *Fr. 2* was also separated by CC (SiO<sub>2</sub>, CHCl<sub>3</sub>/i-PrOH 20:1  $\rightarrow$  10:1): **6** (230 mg) and **7** (34 mg). *Fr. 3* was subjected to CC (*MCI CHP20P* gel, MeOH/H<sub>2</sub>O 0:10  $\rightarrow$  4:6): **8** (2.1 g) and **4** (3.2 g).

*Tinocapilactone A* (=rel-(2Z)-2-[(1E)-2-[1R,4R,4aR,7R,8S,8aS]-3,4,4a,5,6,7,8,8a-Octahydro-4-hydroxy-7-(methoxycarbonyl)-4a,8-dimethyl-3-oxo-1,4-etheno-1H-2-benzopyran-8-yl]ethenyl]but-2-enedioic Acid Dimethyl Ester; **1**): Colorless amorphous powder. UV (CHCl<sub>3</sub>): 264 (4.09).  $[\alpha]_D^{20} = +34.9$  ( $c = 0.2$ , CHCl<sub>3</sub>). IR (KBr): 3482 (OH), 2954, 1746 (C=O), 1716 (C=O), 1437, 1375, 1268, 1205, 1154, 927, 751. <sup>1</sup>H- and <sup>13</sup>C-NMR: *Table*. ESI-MS (pos.): 449 ([*M* + H]<sup>+</sup>). HR-ESI-MS: 449.1815 ([*M* + H]<sup>+</sup>, C<sub>23</sub>H<sub>29</sub>O<sub>7</sub><sup>+</sup>; calc. 449.1812).

*Tinocapilactone B* (=rel-(1R,4R,4aR,7R,8S,8aS)-8-[(1E)-2-(2,5-Dihydro-2-methoxy-5-oxofuran-3-yl)ethenyl]-3,4,4a,5,6,7,8,8a-octahydro-4-hydroxy-4a,8-dimethyl-3-oxo-1,4-etheno-1H-2-benzopyran-7-carboxylic Acid Methyl Ester; **2**): Colorless amorphous powder. UV (CHCl<sub>3</sub>): 262 (4.11).  $[\alpha]_D^{20} = +57.3$  ( $c = 0.6$ , CHCl<sub>3</sub>). IR (KBr): 3466 (OH), 2922, 2851, 1761 (C=O), 1728 (C=O), 1464, 1374, 1154, 972, 723. <sup>1</sup>H- and <sup>13</sup>C-NMR: *Table*. ESI-MS (pos.): 419 ([*M* + H]<sup>+</sup>). HR-ESI-MS: 419.1710 ([*M* + H]<sup>+</sup>, C<sub>22</sub>H<sub>27</sub>O<sub>7</sub><sup>+</sup>; calc. 419.1706).

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